

Large-Scale Nucleotide Sequence Alignment for Universal Screening PCR Assay Design

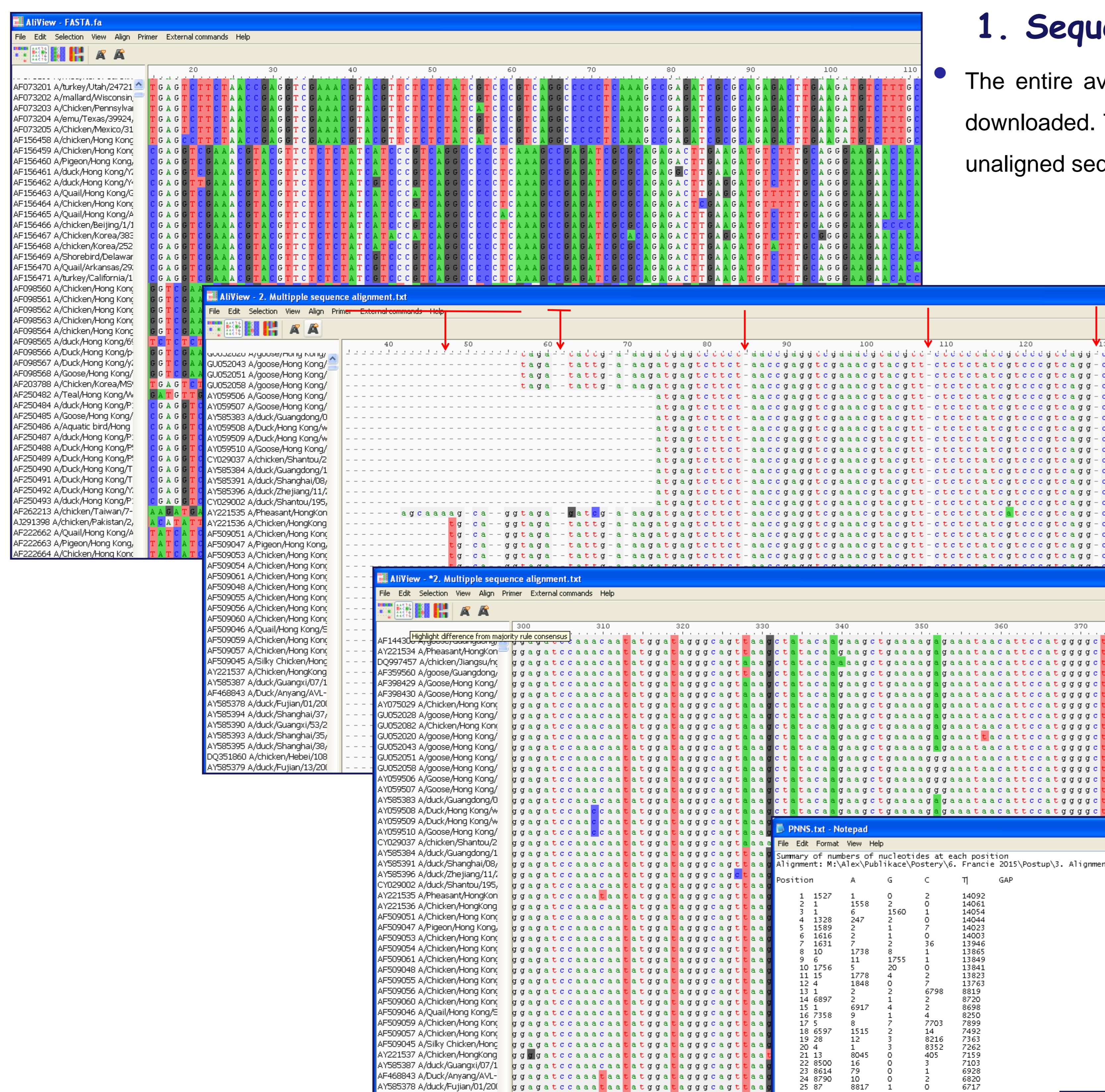
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Introduction

An essential feature of each diagnostic PCR or qPCR assay is to detect as many genetic variants of a given virus or bacterial taxon as possible. A crucial step in such a screening assay design is a proper oligonucleotide selection. Therefore, identification of the evolutionarily most conserved regions of the targeted virus genome is of utmost importance. However, this crucial step is frequently underestimated in the literature. We set up a simple workflow how to align, process, and evaluate a huge set of homologous nucleotide sequences in order to reveal the variability at each particular position. All of the steps were performed by using an ordinary desktop computer without the need for extensive mathematical or computational skills. The workflow was demonstrated on aligning more than 15,000 avian influenza A virus (IAV) M segment nucleotide sequences. The whole procedure takes a few hours and usually do not exceed a half working day in dependence on the internet connection speed and server occupancy respectively.



1. Sequence download

- The entire avian IAV M segment nucleotide sequence content of the Influenza Virus Sequence Database [1] was downloaded. The data contained 15,622 sequences (14-Jul-2015) with the corresponding FASTA file of 15,5 Mb. The unaligned sequences were shown by using the AliView program [2].

2. Multiple sequence alignment (MSA)

- For MSA the online version of the MAFFT v. 7 (Multiple Alignment using Fast Fourier Transform [3]). was implemented. The sequences were compared relative to the reference sequence.
- The figure illustrates a row alignment by using the AliView software [2].

3. Alignment validation

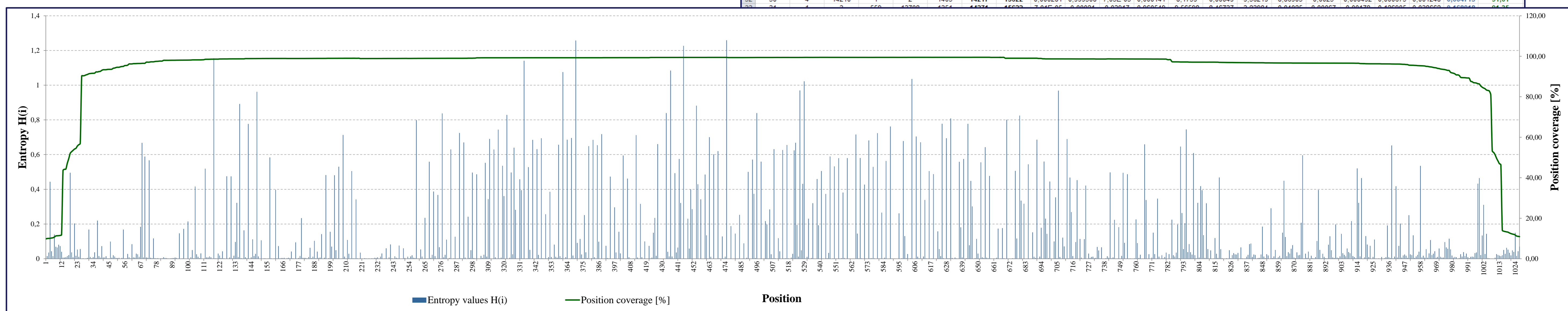
- Preparation of the final alignment requires alignment processing: (editing, trimming, removal of non informative, short, and low quality sequences, and restoration of the alignment continuity (Fig. 2 red arrows). This step was performed in AliView software [2].

4. Positional nucleotide numerical summary (PNNS) calculation

- PNNS - counting the exact number of informative positions (i.e., A, T(U), C, G and gaps) in each column of the alignment by using the BioEdit v.7 program [4].

5. Estimation of sequence variability

- Next, the PNNS was converted to entropy values. The entropy quantifies the amount of variability in each column of the alignment. In this step the EntropyCalculator software [5] was implemented.
- The figure shows the EntropyCalculator output which includes the original PNNS (columns A-F), sum of the informative positions (column G), the entropy calculation chain (columns I-T) as well as the entropy and % coverage values (columns U and V).



6. Results

- Finally, the entropy and %coverage values (Fig. 5, columns U and V) were summarised in a chart where the entropy was drawn as column diagram (blue) related to the main y axis while the % coverage was shown as line graph (green) related to the secondary y axis.
- The graph clearly shows the presence and distribution of the most variable (the highest columns) and also the conserved positions (Hi converging to zero) suitable for oligonucleotide selection.

References

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